Supporting Information

Role of nitrate in conditioning aquifer sediments for technetium bioreduction

Gareth T W Law[†], Andrea Geissler[‡], Christopher Boothman[‡], Ian T Burke[†], Francis R Livens^{‡§}, Jonathan R Lloyd[‡], Katherine Morris^{†*}

 [†]Earth System Science Institute, School of Earth and Environment, University of Leeds, Leeds, LS2 9JT, U.K.
[‡]Williamson Research Centre for Molecular Environmental Science, School of Earth, Atmospheric and Environmental Sciences, The University of Manchester, Manchester, M13 9PL, U.K.
[§]Centre for Radiochemistry Research, and School of Earth, Atmospheric and Environmental Sciences, The University of Manchester, M13 9PL, U.K.

Number of pages - 25 Number of figures - 2 Number of tables - 14

Table of Contents

1.1 Sample collection and storage	S3
1.2. Geochemical methods	S3
1.3. XAS sample preparation	S3
1.4. XAS analysis	S4
1.5. PCR analysis	S4
1.6. References	S24

Figures and Tables

Figure 1. Eh incubation	n time series data	S6
Figure 2. Tc EXAFS at	nd associated Fourier transforms	

* *Corresponding author: School of Earth and Environment, The University of Leeds, Leeds, LS2 9JT, U.K.

Tel: +44 131 343 6723

Fax: +44 113 343 6716

Email address: k.morris@leeds.ac.uk

Table 1. Synthetic groundwater base composition	
Table 2. Synthetic groundwater media compositions	S8
Table 3. Sediment elemental composition	S9
Table 4. EXAFS modeling results	S10
Table 5. Carbonate buffered 16S RNA gene sequences at 0 days	S11
Table 6. High nitrate 16S RNA gene sequences at 0 days	S13
Table 7. Carbonate buffered 16S RNA gene sequences at 24 days	S14
Table 8. Carbonate buffered 16S RNA gene sequences at 250 days	S15
Table 9. High nitrate 16S RNA gene sequences at 30 days	S17
Table 10. High nitrate 16S RNA gene sequences at 60 days	S18
Table 11. High nitrate 16S RNA gene sequences at 80 days	S19
Table 12. High nitrate NarG sequences at 0 days	S20
Table 13. High nitrate NarG sequences at 30 days	S22
Table 14. Quantity of PCR determined Geobacteraceae genomic DNA	S24

1.1. Sample collection and Storage

Sediments, upon collection, were transferred directly into sterile HDPE sample containers, sealed, and stored at 5 °C in darkness. Experiments then began within one month of sediment sampling.

1.2. Geochemical methods

Carbonate-free total sediment organic carbon content was measured by combustion according to the method of Verardo et al., (1990). Total dissolved porewater Fe was measured photometrically using the Ferrozine test (Stookey, 1970), according to the method of Viollier et al., (2000). Porewater manganese was measured photometrically with the PAN indicator method (Brewer and Spencer, 1971). Porewater nitrite was measured photometrically using the method of Harris and Mortimer, (2002).

1.3. XAS Sample Preparation

After 10 days of incubation, pre-reduced sediment microcosms were geochemically monitored prior to XAS sampling. Approximately 40 μ M and 70 μ M of added Tc (ca. 325 μ M) remained in solution in the carbonate buffered and high-nitrate systems, respectively. After geochemical analysis, sediment slurries were centrifuged (12 minutes, 2000 g) and approximately 0.5 g of sediment (water content < 50 %) was mounted for XAS analysis on an air-tight XAS Perspex sample cell with Kapton windows. The sample cell was then double contained in heat sealed plastic bags and stored at -80 °C under argon until analysis. All sample manipulations were conducted in an O₂-free atmosphere.

1.4. XAS Analysis

Technetium K-edge spectra were collected in fluorescence mode on a 30 element solid-state Ge detector on the ultra dilute spectroscopy line (16.5) at the UK CLRC Daresbury SRS, using focusing optics and a Si-220 double crystal monochromator, calibrated from the K-edge of a Mo foil. A total of 10-12 scans were collected for each sample and averaged to improve the signal-to-noise ratio. The EXAFS spectra were background subtracted using EXPLINE and analyzed with EXCURV98 using full curved wave theory (Gurman et al., 1995). Phase shifts were determined from ab initio calculations using Hedin-Lundqvist potentials and von Barth ground states (Binsted, 1998). The data were fitted in k^3 space by defining a theoretical model which was informed by the relevant literature (Wharton et al., 2000; Lukens et al., 2002; Maes et al., 2004; Morris et al., 2008) and used whole integer values for shell of back-scatters around the central atom. Shells of backscatter atoms were added around a central ⁹⁹Tc atom and by refining an energy correction Ef (the fermi energy), the absorber-scatter distance, the number of atoms in each shell, and the least square residual (the R factor; Binsted, 1992) was minimized (Morris et al., 2008). Shells were only included in the model fit if the overall R-factor was improved by > 5 %.

1.5. PCR analysis

A standard curve for the qPCR reaction was created by plotting the cycle threshold values of the qPCR performed on a dilution series of DNA obtained from *Geobacter sulfurreducens* (GenBank Accession Number U13928) against the log of DNA template concentration. Dilution series concentrations ranging from 0.1 ng μ l⁻¹ to 0.88 pg μ l⁻¹,

were analysed in triplicate and had an R^2 value of 0.98. PCR amplification was performed in triplicate on a Stratagene MX3000P qPCR machine using 0.25 μ M of each primer and qPCR SYBR Green Master Mix (Stratagene) with an initial step of 94 °C for 10 minutes followed by 40 cycles of 94 °C for 30 seconds, 58 °C for 30 seconds and 72 °C for 30 seconds. A dissociation curve was run between 94 °C and 58 °C to check primer specificity. Cycle threshold was determined automatically.



Figure 1. Microbially active microcosm Eh incubation time-series data (day 0-250). Dark blue circles = carbonate buffered system, Green squares = unamended system, Red diamonds = low-nitrate system, Light blue inverted triangles = high-nitrate system. Error bars represent 1 σ experimental uncertainty from triplicate microcosm experiments (where not visible, error bars are within the symbol size). Eh did not change in sterile controls.

Carbonate buffered



Figure 2. Background subtracted, normalized, and k^3 weighted ⁹⁹Tc K edge EXAFS spectra (left) and corresponding Fourier transforms (right) for reduced sediments from the carbonate buffered and high nitrate systems (black) and for the best modeled fit obtained in supporting information Table 3 (red).

Compound	g L-1
KCl	0.0066
MgSO ₄ .7H ₂ O	0.0976
MgCl ₂ .6H ₂ O	0.0810
Na ₂ SiO ₃	0.0829
NaCl	0.0094

Table 1. Synthetic groundwater media base composition (Wilkins et al., 2007).

Table 2. Synthetic groundwater media compositions.

Microcosm	Base media	$NO_3^-(mM)$	SO_4^{2-} (mM)	Na-acetate (mM)	рН
Unamended	Wilkins et al, 2007	0.3	0.4	10.0	5.5
Low-nitrate	Wilkins et al, 2007	2.0	0.4	10.0	5.5
High-nitrate	Wilkins et al, 2007	10.0	0.4	10.0	5.5
Carbonate buffered*	Wilkins et al, 2007	0.3	0.4	10.0	7.0

* 3 mM NaHCO₃ and 1.7 mM CaCO3 were added to the carbonate buffered groundwater media.

Element	Concentration
Si	35 wt %
Al	5.8 wt %
Fe	3.1 wt %
Κ	2.7 wt %
Na	1.0 wt %
Mg	0.5 wt %
Ti	0.4 wt %
Ca	0.2 wt %
Ba	0.2 wt %
Mn	0.1 wt %
Р	0.02 wt %
Ba	0.2 wt %
Zr	251 ppm
Rb	119 ppm
V	81 ppm
Sr	58 ppm
Zn	51 ppm
Pb	42 ppm
Cr	30 ppm
Cu	<30 ppm
Nd	21 ppm
La	18 ppm
Ni	17 ppm
Со	<10 ppm
LOI	4.07 wt %

Table 3. Sediment major and minor elemental composition

Sample	Shell	О-Т	r (Å)	$2\sigma^2$ (Å ²)	R
Carbonate buffered	1	6-0	1.99	0.016	42
	1	6-O	2.01	0.016	29
	2	1-1c	2.52	0.014	
	1	6-O	2.00	0.016	32
	2	1-Fe	2.69	0.008	
High-nitrate	1	6-0	2.00	0.018	42
	1	6-O	2.00	0.016	28
	2	1-Tc	2.53	0.011	
	1	6-O	2.00	0.016	34
	2	1-Fe	2.69	0.008	

Table 4. Summary of EXAFS modelling results for reduced sediments representative ofthe Sellafield site.

O-T is the modelled occupancy and type, r (Å) is the interatomic distance, $2\sigma^2$ (Å²) is the Debye-Waller factor, and R is the least squares residual.

Clone number	Accesion number	Closest relative (accession number)	Similarity (% Match)	% present in clone library	Phylogenetic Division
AT0-1	GQ903133	Uncultured bacterium	409/443	2.3	Alphaproteobacteria
		TSC131 (EU360041)	(92%)	(1/43)	r T T
AT0-2	GQ903134	Uncultured bacterium	400/425	6.97	Acidobacteria
		5V79 (DQ663849)	(94%)	(3/43)	
AT0-3	GQ903135	Uncultured Nitrosovibrio	387/395	4.65	Unknown
		sp. B4_14 (AM773598)	(97%)	(2/43)	
AT0-5	GQ903136	Uncultured bacterium	358/395	4.65	Acidobacteria
		AH31 (AY963324)	(90%)	(2/43)	
AT0-7	GQ903137	Uncultured bacterium	381/399	9.3	Acidobacteria
		GASP-MA3W3_H06 (EF663817)	(95%)	(4/43)	
AT0-10	GQ903138	Uncultured Planctomyces	330/356	2.3	Planctomycetes
		sp. BD1-23 (AB015527)	(92%)	(1/43)	
AT0-11	GQ903139	Uncultured bacterium	320/352	2.3	Betaproteobacteria
		SX2-19 (DQ469209)	(90%)	(1/43)	
AT0-14	GQ903140	Uncultured bacterium	386/398	2.3	Acidobacteria
		C33.24SM (AF432814)	(96%)	(1/43)	
AT0-15	GQ903141	Uncultured bacterium	324/360	2.3	Unknown
		S15-3 (EU680463)	(90%)	(1/43)	
AT0-17	GQ903142	Uncultured alpha proteo-	409/415	6.97	Alphaproteobacteria
		bacterium GASP-45KA- 43-C06 (EU044312)	(98%)	(3/43)	
AT0-18	GQ903143	Unidentified bacterium	471/504	4.65	Acidobacteria
	-	H02_SGPO01(EF221060)	(93%)	(2/43)	
AT0-19	GQ903144	Uncultured bacterium	403/433	4.65	Unknown
		isolate High.60.F3.HB13 (DQ195631)	(93%)	(2/43)	
AT0-20	GQ903145	Uncultured Verruco-	451/474	2.3	Verrucomicrobia
		microbia bacterium A07 WMSP1 (DQ450782)	(95%)	(1/43)	
AT0-21	GQ903146	Uncultured bacterium	330/361	2.3	Unknown
		anNA12 (EF034689)	(91%)	(1/43)	
AT0-24	GQ903147	Uncultured bacterium	402/416	2.3	Acidobacteria
		Amb_16S_1165 (EF018714)	(96%)	(1/43)	
AT0-25	GQ903148	Uncultured bacterium	378/420	4.65	Acidobacteria
		FCPU671 (EF516884)	(90%)	(2/43)	
AT0-26	GQ903149	Uncultured Acidobacteria	345/382	2.3	Acidobacteria
		bacterium WSD-032 (EF588358)	(90%)	(1/43)	
AT0-28	GQ903150	Uncultured bacterium	516/523	2.3	Unknown
		FAC55 (DQ451494)	(98%)	(1/43)	
AT0-29	GQ903151	Uncultured bacterium	379/393	6.97	Acidobacteria
		JG30-KF-C37(AJ536864)	(96%)	(3/43)	
AT0-30	GQ903152	Uncultured bacterium	494/497	2.3	Betaproteobacteria
		S15-143 (EU680411)	(99%)	(1/43)	
AT0-32	GQ903153	Uncultured bacterium	393/399	4.65	Acidobacteria
		GASP-16KA-214-F03 (EU043920)	(98%)	(2/43)	

Table 5. Phylogenetic affiliation of 16S rRNA gene sequences detected in the clone library from the carbonate buffered system at 0 days.

AT0-33	GQ903154	Uncultured bacterium 3-	488/497	2.3	Unknown
		33 (EU637706)	(98%)	(1/43)	
AT0-34	GQ903155	Uncultured Verruco-	478/524	2.3	Verrucomicrobia
		microbia bacterium A07 WMSP1 (DQ450782)	(91%)	(1/43)	
AT0-37	GQ903156	Uncultured gamma	466/498	4.65	Unknown
		proteo-bacterium	(93%)	(2/43)	
		N41.113PG (AF431365)			
AT0-43	GQ903157	Variovorax sp. KS2D-23	489/498	2.3	Betaproteobacteria
		(AB196432)	(98%)	(1/43)	-
AT0-50	GQ903158	Uncultured bacterium	455/461	2.3	Acidobacteria
		FCPP450 (EF516894)	(98%)	(1/43)	
AT0-53	GQ903159	Uncultured Verruco-	421/451	2.3	Verrucomicrobia
		microbiales bacterium	(93%)	(1/43)	
		AhedenT24 (FJ475385)			

Clone	Accesion	Closest relative	Similarity	% present	Phylogenetic
name	number	(accession number)	(% Match)	in clone	Division
		× /	· · · · ·	library	
CT0-1	GQ903176	Uncultured bacterium	485/506	2.0	Verrucomicrobia
	-	FCPT606 (EF516172)	(95%)	(1/51)	
CT0-2	GQ903177	Uncultured alpha	417/445	21.6	Alphaproteobacteria
	-	proteobacterium glen99 6	(93%)	(11/51)	
		(AY150881)	· · ·		
СТ0-3,	GQ903178	Uncultured bacterium 9S26	479/480	15.7	Acidobacteria
CT0-55		(DQ664128)	(99%)	(8/51)	
CT0-5	GQ903179	Uncultured bacterium	468/492	2.0	Unknown
		FCPP437 (EF516340)	(95%)	(1/51)	
CT0-6	GQ903180	Uncultured bacterium FAC71	489/492	2.0	Acidobacteria
		(DQ451510)	(99%)	(1/51)	
CT0-8	GQ903181	Uncultured bacterium	464/470	3.9	Unknown
		S22.41WL (AF432729)	(98%)	(2/51)	
CT0-9	GQ903182	Unidentified bacterium	451/480	3.9	Unknown
		D01_SGPO01 (EF220972)	(93%)	(2/51)	
CT0-14	GQ903183	Uncultured bacterium	438/448	3.9	TM7
	-	DOK CONFYM clone792	(97%)	(2/51)	
		(DQ828993)			
CT0-16	GQ903184	Uncultured alpha proteo-	431/438	7.8	Alphaproteobacteria
	-	bacterium AKYG981	(98%)	(4/51)	
		(AY921934)	· · ·		
CT0-17	GQ903185	Uncultured forest soil	470/473	7.8	Acidobacteria
		bacterium DUNssu227	(99%)	(4/51)	
		(AY913429)			
CT0-18	GQ903186	Uncultured bacterium 38	475/484	3.9	Acidobacteria
		(EF474280)	(98%)	(2/51)	
CT0-20	GQ903187	Unidentified bacterium AI-	454/470	11.8	Unknown
		2M_E11 (EF219790)	(96%)	(6/51)	
CT0-28	GQ903188	Uncultured bacterium AS51	459/463	2.0	Acidobacteria
		(AY963415)	(99%)	(1/51)	
CT0-29	GQ903189	Uncultured bacterium	461/493	2.0	Bacteroidetes
		FFCH12595 (EU133712)	(93%)	(1/51)	
CT0-31	GQ903190	Uncultured bacterium	443/451	2.0	Alphaproteobacteria
		DOK_CONFYM_clone440	(98%)	(1/51)	
		(DQ828690)			
CT0-40	GQ903191	Uncultured bacterium	450/452	2.0	Unknown
		DUNssu007(+1B)(OTU#078)	(99%)	(1/51)	
		(AY724094)			
CT0-43	GQ903192	Uncultured proteobacterium	469/496	2.0	Unknown
		TSC21 (EU359932)	(94%)	(1/51)	
CT0-51	GQ903193	Uncultured Acidobacteria	459/485	2.0	Acidobacteria
		bacterium WSD-032	(94%)	(1/51)	
		(EF588358)			
CT0-53	GQ903194	Uncultured bacterium	446/453	2.0	Acidobacteria
		FCPP453 (EF516137)	(98%)	(1/51)	

Table 6. Phylogenetic affiliation of 16S rRNA gene sequences detected in 16S rRNA clone library from the high nitrate system at 0 days.

Clone	Accession	Closest relative	Similarity	% present	Phylogenetic
name	number	(accession number)	(%	in clone	Division
			Match)	library	
AT24-1	GQ903160	Sphingomonas sp. EC-K085	346/361	4.5	Alphaproteobacteria
		(AB264174)	(95%)	(2/44)	
AT24-2	GQ903161	Uncultured bacterium RS 8-	476/494	13.6	Betaproteobacteria
		Bact23 (AJ867667)	(96%)	(6/44)	
		Janthinobacterium lividum	469/489		
		GA01 (DQ473538)	(95%)		
AT24-3	GQ903162	Uncultured bacterium	466/485	2.3	Unknown
		Elev_16S_1406 (EF020015)	(96%)	(1/44)	
AT24-4	GQ903163	Uncultured candidate	444/446	2.3	TM7
		division TM7 bacterium AI- 2M C01 (EF219756)	(99%)	(1/44)	
AT24-5	GQ903164	Burkholderia sediminicola	476/482	6.8	Betaproteobacteria
		strain HU2-65W	(98%)	(3/44)	1
		(EU035613)			
AT24-6	GQ903165	Uncultured gamma	483/488	6.8	Gammaproteo-
		proteobacterium S-A1	(98%)	(3/44)	bacteria
		(AY622264)			
AT24-8	GQ903166	Uncultured bacterium	477/488	25.0	Betaproteobacteria
		M14C24 (EU331388)	(97%)	(11/44)	
AT24-12	GQ903167	Uncultured Bacteroidetes	480/483	4.5	Bacteroidetes
	-	bacterium CrystalBog5D8 (AY792301)	(99%)	(2/44)	
AT24-13	GO903168	Uncultured soil bacterium	439/458	2.3	Betaproteobacteria
	- (,	L1A.13F01 (AY989483)	(95%)	(1/44)	r
AT24-14	GO903169	Uncultured bacterium	469/492	11.4	Bacteroidetes
		KuyT-ice-74 (EU263768)	(95%)	(5/44)	
AT24-18	GO903170	Unidentified bacterium	464/481	2.3	Acidobacteria
	τ.	C05 SGPL01 (EF221128)	(96%)	(1/44)	
AT24-19	GO903171	Desulfosporosinus sp. A10	499/514	6.8	Firmicutes
		(AJ582756)	(97%)	(3/44)	
AT24-25	GO903172	Uncultured Acidobacteria	471/481	2.3	Acidobacteria
		bacterium WSD-037	(97%)	(1/44)	
		(EF588363)			
AT24-27	GO903173	Uncultured bacterium	448/474	4.5	Betaproteobacteria
		BANW605 (DO264563)	(94%)	(2/44)	T
AT24-31	GQ903174	Dyella koreensis strain BB4	475/489	2.3	Gammaproteo-
		(ÅY884571)	(97%)	(1/44)	bacteria
AT24-32	GQ903175	Ralstonia sp. PHD-12	463/482	2.3	Betaproteobacteria
		(DQ374437)	(96%)	(1/44)	T

Table 7. Phylogenetic affiliation of 16S rRNA gene sequences detected in 16S rRNA clone library from the carbonate buffered system at 24 days.

Clone	Accession	Closest relative	Similarity	%	Phylogenetic
name	number	(accession Number)	(% Match)	present	division
				in clone	
				library	
AT250-1	GQ903237	Geobacter psychrophilus	496/516	2.0	Deltaproteobacteria
		P35 (AY653549)	(96%)	(1/49)	
AT250-2	GQ903238	Uncultured Geobacter sp.	497/515	8.2	Deltaproteobacteria
		NO3_C-10 (AY752772)	(96%)	(4/49)	
AT250-4,	GQ903239	Uncultured bacterium	503/515	8.2	Bacteroidetes
AT250-9	GQ903240	t30d34L66 (FM956250)	(97%)	(4/49)	
AT250-5	GQ903241	Uncultured bacterium 9S26	499/500	4.1	Acidobacteria
	~~~~	(DQ664128)	(99%)	(2/49)	
AT250-6	GQ903242	Uncultured bacterium	492/503	10.2	Betaproteobacteria
	~~~~	M14C24 (EU331388)	(97%)	(5/49)	
AT250-10	GQ903243	Uncultured bacterium	477/498	4.1	Bacteroidetes
	G. 0. 0. 0. 0. 0. 0. 0. 0. 0. 0. 0. 0. 0.	FRC-A2_/35 (EF508009)	(95%)	(2/49)	T T 1
A1250-14	GQ903244	Uncultured bacterium 9854	487/507	4.1	Unknown
A TTO 50 10	G0000045	(DQ664146)	(96%)	(2/49)	
A1250-19	GQ903245	Uncultured bacterium	462/465	2.0	Acidobacteria
4 7 2 5 0 2 0	00000046	BS6/ (AY963503)	(99%)	(1/49)	A *11 / *
A1250-20	GQ903246	Uncultured bacterium	492/509	2.0	Acidobacteria
4 7 2 5 0 2 2	00000047	RB355 (AB240350)	(96%)	(1/49)	TT 1
A1250-22	GQ903247	Uncultured bacterium	480/499	2.0	Unknown
A T 2 5 0 2 2	CO002248	LWS-14642 (EU546377)	(96%)	(1/49)	Destantilates
AT250-23,	GQ903248	Uncultured Bacteroldetes	4/6/483	16.3	Bacteroidetes
A1250-45	GQ903249	(DO501208)	(98%)	(8/49)	
AT250 25	CO002250	(DQ301308)	192/105	2.0	Destaraidatas
A1230-23	GQ903230	I WS DSC 2022	482/493	(1/40)	Bacteroideles
		E W S - KSO - 5952 (EU546744)	(9770)	(1/49)	
AT250-26	GO003251	(E0340744) Uncultured bacterium	/00/515	4.1	Deltanroteobacteria
A1230-20	00903231	$FRC_{-}A2_{-}251$ (FF507072)	(96%)	(2/40)	Denaproteobacterra
AT250-31	GO903252	Uncultured Xylophilus sp	490/499	(2/4)	Retanroteobacteria
111250 51	92705252	CMIB6 (AM936163)	(98%)	(2/49)	Detaproteobacteria
AT250-35	GO903253	Uncultured Verruco-	399/425	20	Verrucomicrobia
111200 00	000200	microbia bacterium	(94%)	(1/49)	· en aconnero ena
		HF032 (EF417621)	(31,0)	(1,1))	
AT250-37	GO903254	Uncultured bacterium	433/453	2.0	Bacteroidetes
		D41S30C27 (FM956915)	(95%)	(1/49)	
AT250-38	GO903255	Uncultured bacterium	321/336	4.1	Bacteroidetes
		D242 27F BAC2 013	(95%)	(2/49)	
		(AB447719)	× ,		
AT250-41	GQ903256	Uncultured bacterium	399/418	2.0	Unknown
	-	Roth04 (AY395221)	(95%)	(1/49)	
AT250-42	GQ903257	Uncultured bacterium	399/423	2.0	Deltaproteobacteria
		LD_Ba_Top_B42	(94%)	(1/49)	
		(EU644267)			
AT250-43	GQ903258	Uncultured bacterium	403/434	2.0	BRC1
		D41S30C84 (FM956960)	(92%)	(1/49)	
AT250-44	GQ903259	Uncultured bacterium	264/265	2.0	Acidobacteria
		FAC71(DQ451510)	(99%)	(1/49)	

Table 8. Phylogenetic affiliation of 16S rRNA gene sequences detected in 16S rRNA clone library from the carbonate buffered system at 250 days.

AT250-47	GQ903260	Uncultured bacterium X9Ba78 (AY607208)	241/260 (92%)	4.1 (2/49)	Chloroflexi
AT250-48	GQ903261	Uncultured bacterium FW42 (F524019)	374/386 (96%)	2.0 (1/49)	Chloroflexi
AT250-50	GQ903262	Uncultured delta proteo- bacterium C32.51SM (AF431389)	214/221 (96%)	2.0 (1/49)	Deltaproteobacteria
AT250-53	GQ903263	Uncultured proteo- bacterium AhedenR37 (FJ475537)	226/227 (99%)	2.0 (1/49)	Unknown

Clone Accession Close		Closest relative	Similarity	%	Phylogenetic	
name	number	(accession number)	(%	present	division	
			Match)	in clone		
			,	library		
CT30-1	GQ903195	Uncultured forest soil	450/455	1.9	Alphaproteobacteria	
		bacterium DUNssu011	(98%)	(1/52)		
		(AY913232)				
CT30-2	GQ903196	Unidentified bacterium	371/374	1.9	Acidobacteria	
		C05_SGPL01 (EF221128)	(99%)	(1/52)		
CT30-3	GQ903197	Uncultured Acidobacteria	326/340	1.9	Acidobacteria	
		bacterium B08_WMSP1 (DQ450707)	(95%)	(1/52)		
CT30-4	GQ903198	Rhodanobacter terrae GP18-1	409/417	1.9	Gammaproteo-	
		(EF166076)	(98%)	(1/52)	bacteria	
CT30-5	GQ903199	Uncultured bacterium	347/385	1.9	Firmicutes	
		MBFOS-11 (EU369170)	(90%)	(1/52)		
CT30-6	GQ903200	Herbaspirillum sp. P-64	290/302	28.8	Betaproteobacteria	
CT30-53	GQ903201	(AM411937)	(96%)	(15/52)		
		Herbaspirillum sp. PIV-34-1	288/302			
		(AJ505863)	(95%)			
CT30-7	GQ903202	Uncultured soil bacterium	318/329	1.9	Acidobacteria	
		CWT ST03_F05 (DQ129198)	(96%)	(1/52)		
CT30-8	GQ903203	Bacillus senegalensis RN2	284/295	13.5	Firmicutes	
		(AB110415)	(96%)	(7/52)		
CT30-9	GQ903204	Burkholderia sp. SB5	337/337(1	1.9	Betaproteobacteria	
		(AJ971354)	00%)	(1/52)		
CT30-10	GQ903205	Uncultured Acidobacteria	373/376	1.9	Acidobacteria	
		bacterium WSD-032 (EF588358)	(99%)	(1/52)		
CT30-11	GQ903206	Uncultured bacterium MSC49	289/294	3.8	Betaproteobacteria	
	-	(AJ830735)	(98%)	(2/52)	1	
CT30-13	GQ903207	Uncultured bacterium M40C1	493/502	1.9	Betaproteobacteria	
	2	(EU331385)	(98%)	(1/52)	1	
CT30-14	GQ903208	Uncultured bacterium	405/409	1.9	Acidobacteria	
		FCPT732 (EF516752)	(99%)	(1/52)		
CT30-15	GQ903209	Sporolactobacillus dextrus	334/345	11.5	Firmicutes	
CT30-47	GQ903210	IAM 12380 (D16282)	(96%)	(6/52)		
CT30-17	GQ903211	Uncultured bacterium PA2	388/393	11.5	Firmicutes	
		01_D06 (AM900227)	(98%)	(6/52)		
CT30-23	GQ903212	Uncultured bacterium GPT3	381/388	3.8	Gammaproteo-	
		(AY706437)	(98%)	(2/52)	bacteria	
CT30-27	GQ903213	Uncultured forest soil	478/481	1.9	Acidobacteria	
		bacterium DUNssu072	(99%)	(1/52)		
		(AY913295)				
CT30-32	GQ903214	Uncultured bacterium 9S26	491/495	1.9	Acidobacteria	
		(DQ664128)	(99%)	(1/52)		
CT30-33	GQ903215	Uncultured Bacilli bacterium	400/420	3.8	Firmicutes	
CT30-42	GQ903216	X3Ba61 (AY607149)	(95%)	(2/52)		

Table 9. Phylogenetic affiliation of 16S rRNA gene sequences detected in 16S rRNA clone library from the high nitrate system at 30 days.

Clone	Accession	Closest relative	Similarity	%	Phylogenetic	
name	number	iber (accession number)		present in clone library	division	
CT60-1	GQ903217	Uncultured beta proteo-	442/478	2.1	Betaproteobacteria	
		(EF417722)	(92%)	(1/48)		
CT60-2	GQ903218	Uncultured bacterium PA2	384/395	8.3	Firmicutes	
		01_D06 (AM900227)	(97%)	(4/48)		
CT60-3	GQ903219	Sporolactobacillus dextrus	343/356	8.3	Firmicutes	
		IAM 12380(D16282)	(96%)	(4/48)		
CT60-4	GQ903220	Uncultured bacterium	468/487	2.1	Betaproteobacteria	
		M40C1 (EU331385)	(96%)	(1/48)		
CT60-5	GQ903221	Uncultured beta proteo-	365/385	60.4	Betaproteobacteria	
		bacterium JG35+U3-JT43 (AM084888)	(94%)	(29/48)	-	
CT60-16	GO903222	Uncultured soil bacterium	492/537	4.2	TM7	
		MJK38 (EF540381)	(91%)	(2/48)		
CT60-20	GO903223	Uncultured forest soil	432/435	2.1	Acidobacteria	
		bacterium DUNssu174 (AY913381)	(99%)	(1/48)		
CT60-27	GO903224	Uncultured bacterium	399/418	2.1	Gammaproteo-	
		1/5/6F (FJ380139)	(95%)	(1/48)	bacteria	
CT60-31	GO903225	Uncultured Bacillus sp.	304/327	4.2	Firmicutes	
СТ60-32	GO903226	PmeaMucB8 (EU249961)	(92%)	(2/48)		
CT60-45	GO903227	Uncultured bacterium LS4-	399/424	4.2	Gammaproteo-	
	X · · · · · ·	200 (AB234278)	(94%)	(2/48)	bacteria	
CT60-49	GO903228	Uncultured Rhodanobacter	358/389	2.1	Gammaproteo-	
		sp. GASP-WC1S1_C08 (EF074391)	(92%)	(1/48)	bacteria	

Table 10. Phylogenetic affiliation 16S rRNA gene sequences detected in 16S rRNA clone library from the high nitrate system at 60 days.

Clone	Accession	Closest relative	Similarity	% present	Phylogenetic division	
паше	number	(accession number)	(70 Match)	library		
CT80-1	GQ903229	Uncultured Geobacteraceae	499/515	28.3	Deltaproteobacteria	
		bacterium M16_6691 (EF669008)	(96%)	(13/46)		
CT80-2	GQ903230	Herbaspirillum sp.	473/484	58.7	Betaproteobacteria	
		PIV-34-1 (AJ505863)	(97%)	(27/46)	-	
CT80-3	GQ903231	Mucilaginibacter sp. BR5-	464/484	2.2	Bacteroidetes	
		28 (EU423305)	(95%)	(1/48)		
CT80-8	GQ903232	Uncultured Acidobacteria	482/491	2.2	Acidobacteria	
		bacterium WSD-032 (EF588358)	(98%)	(1/48)		
CT80-9	GQ903233	Uncultured bacterium	480/495	2.2	Firmicutes	
	-	ph5Glu032-5C (AY527739)	(96%)	(1/48)		
CT80-16	GQ903234	Uncultured Geobacteraceae	465/501	2.2	Deltaproteobacteria	
		bacterium M16_6691 (EF669008)	(92%)	(1/48)		
CT80-39	GQ903235	Uncultured bacterium KD9-	475/484	2.2	Bacteroidetes	
		53 (AY218669)	(98%)	(1/48)		
CT80-47	GQ903236	Sporolactobacillus dextrus	485/491	2.2	Firmicutes	
		IAM 12380 (D16282)	(98%)	(1/48)		

Table 11. Phylogenetic affiliation of 16S rRNA gene sequences detected in 16S rRNA clone library from the high nitrate system at 80 days.

Clone name Accession **Closest amino acid sequence** Similarity % (accession number) (% Match) number present CT0-narG 1 FN554828 putative dissimilatory membrane-bound 179/203 40 nitrate reductase [uncultured bacterium] (88.2 %) (AAM62209) clone C71 CT0-narG 3 FN554829 membrane bound nitrate reductase 181/205 2 [uncultured bacterium] (ABF20963) (88.3%) clone LT-250 43 membrane bound nitrate reductase CT0-narG 4 FN554830 179/207 18 [uncultured bacterium] (ABF20963) (86.5 %) clone LT-250 43 respiratory nitrate reductase 1 alpha chain CT0-narG 5 FN554831 160/2082 [Marinobacter sp. ELB17] (ZP 01736799 (76.9%)or EBA00369) CT0-narG 11 FN554832 putative dissimilatory membrane-bound 176/208 2 nitrate reductase [unidentified bacterium] (84.6 %) (AAS59913) clone NCGC34 CT0-narG 12 FN554833 membrane bound nitrate reductase 4 185/204 [unidentified bacterium] (AAY56563) (90.7%) clone 10g44 nitrate reductase 1, alpha subunit 174/204 [Herminiimonas arsenicoxydans] (85.3 %) (CAL61819) CT0-narG 13 FN554834 membrane bound nitrate reductase 157/202 2 [uncultured bacterium](ABF20878) (77.7%) clone UT-075 46 nitrate reductase, alpha subunit 136/202 [Rubrobacter xylanophilus DSM 9941] (67.3%)(YP 643983) CT0-narG 17 2 FN554835 dissimilatory membrane-bound nitrate 188/207 reductase [uncultured bacterium] (90.8%) (CAL80874) clone JG35-2-AG-narG81 nitrate reductase, alpha subunit 177/208 [Thiobacillus denitrificans (85.1%) ATCC 25259] (YP_315161) putative dissimilatory membrane-bound CT0-narG 15 FN554836 182/197 4 nitrate reductase [uncultured bacterium] (92.4%)(AAP79341) clone GA7 CT0-narG 38 putative dissimilatory membrane-bound FN554837 185/200 4 nitrate reductase [uncultured bacterium] (92.5%) (AAM62164) clone A8 CT0-narG 19 FN554838 nitrate reductase [uncultured 168/183 4 bacterium](CAL92067) clone Dbf75 (91.8%) nitrate reductase, alpha subunit 129/182 [Burkholderia cepacia AMMD] (70.8%)(YP 777592) CT0-narG 25 putative dissimilatory membrane-bound 177/209 2 FN554839 nitrate reductase [unidentified bacterium] (84.7%) (AAS59880) clone NCGB54 CT0-narG 32 FN554840 respiratory nitrate reductase alpha chain 4 127/160 (NarG) [Ralstonia eutropha H16] (79.4%)(YP 841777) CT0-narG 33 FN554841 membrane bound nitrate reductase 194/204 2 [unidentified bacterium] (95.1%)(AAY56506) clone 17g20 CT0-narG 34 FN554842 putative dissimilatory membrane-bound 178/208 2 nitrate reductase [unidentified (85.6%)

Table 12. NarG sequences retrieved from the high nitrate system at day 0 and their closest matches in the GenBank.

bacterium](AAS59904) clone NCSF38

CT0-narG_40	FN554843	putative dissimilatory membrane-bound nitrate reductase [uncultured bacterium] (AAX59101) clone GRAMD33	139/207 (67.1%)	2
CT0-narG_44	FN554844	putative dissimilatory membrane-bound nitrate reductase [uncultured bacterium] (AAM62255) clone D101 putative dissimilatory membrane-bound nitrate reductase [uncultured bacterium] (AAX50116) clone GRAM022	190/207 (91.8%)	2
CT0-narG_45	FN554845	putative dissimilatory membrane-bound nitrate reductase [unidentified bacterium] (AAS59880) clone NCGB54	179/207 (86.5%)	2

Clone Accession **Closest amino acid sequence** Similarity % (accession number) (% Match) number present CT30-narG 1 FN554846 nitrate reductase alpha subunit Bacillus ~77% 19 subtilis (ABG33916) CT30-narG 5 FN554847 membrane bound nitrate reductase 200/207 15 [uncultured bacterium] clone LT-075 21 (96.6%) Respiratory nitrate reductase alpha subunit 149/208 [Geobacillus thermodenitrificans NG80-(71.6%)2] (YP 001125821) CT0-narG 2 dissimilatory membrane-bound nitrate FN554848 200/207 11 reductase [uncultured bacterium] clone JG35-(96.6%) K4-AG-narG1 (CAL80836) Respiratory nitrate reductase alpha subunit 145/204 [Geobacillus thermodenitrificans NG80-2] (71.1%)(YP 001125821) CT30-narG 25 FN554849 dissimilatory membrane-bound nitrate 179/197 6 reductase [uncultured bacterium] clone (90.9%) JG35-K4-narG9 (CAL80838) nitrate reductase, alpha subunit [Bacillus 153/200 coagulans 36D1] (ZP 01695255) (76.5%)CT30-narG 4 FN554850 dissimilatory membrane-bound nitrate 175/205 4 reductase [uncultured bacterium] (85.4 %) (AAY56563) clone JG35-K4-AG-narG3 nitrate reductase, alpha subunit [Bacillus 131/206 coagulans 36D1] (ZP 01695255) (63.4%)168/199 CT30-narG 15 FN554851 dissimilatory membrane-bound nitrate 6 reductase [uncultured bacterium] (84.4%) (AAY56563) clone JG35-K4-AG-narG3 nitrate reductase, alpha subunit [Bacillus 123/200 coagulans 36D1] (ZP 01695255) (61.5 %) membrane bound nitrate reductase CT30-narG 11 FN554852 129/207 9 [uncultured bacterium] clone LT-(62.7%) 075 23(ABF20968)

nitrate reductase, alpha subunit [Bacillus

putative dissimilatory membrane-bound

nitrate reductase [uncultured bacterium]

[unidentified bacterium] (AAY56563)

dissimilatory membrane-bound nitrate

(CAL80849) clone JG35+U1-AG-narG21 nitrate reductase 1, alpha subunit

putative dissimilatory membrane-bound

nitrate reductase [unidentified bacterium]

coagulans 36D1] (ZP 01695255)

membrane bound nitrate reductase

reductase [uncultured bacterium]

[Herminiimonas arsenicoxydans]

(AAS59880) clone NCGB54 nitrate reductase, alpha subunit

[Methylobacterium sp. 4-46]

membrane bound nitrate reductase

[uncultured bacterium] (ABF20967)

(AAM62209) clone C71

clone 10g44

(YP 001099944)

(ZP 01851204)

clone LT-075 21

CT30-narG 20

CT30-narG 32

CT30-narG 34

CT30-narG_35

FN554853

FN554854

FN554855

FN554856

129/207

(62.3%)

182/207

(87.9%)

187/204

(91.7%)

180/204

(88.2%)

176/204

(86.3%)

180/207

(87.0%)

171/207

(82.6%)

201/207

(97.1%)

149/208

(71.6%)

9

2

2

2

Table 13. NarG sequences retrieved from the high nitrate system at day 30 and their closest matches in the GenBank.

Respiratory nitrate reductase alpha subunit

[Geobacillus thermodenitrificans NG80-2]

(YP_001125821)

CT30-narG_37	FN554857	membrane bound nitrate reductase [uncultured bacterium] (ABF20963) clone	180/208 (86.5%)	2
		LT-250 43	· · · ·	
CT30-narG 46	FN554858	putative dissimilatory membrane-bound	189/206	2
_		nitrate reductase [uncultured	(91.7%)	
		bacterium](AAM62164) clone A8	. ,	
CT30-narG 47	FN554859	nitrate reductase [uncultured bacterium]	186/205	2
_		(CAL92067)clone DBf75	(90.7%)	
CT30-narG 27	FN554860	dissimilatory membrane-bound nitrate	200/204	2
_		reductase [uncultured bacterium]	(98.0%)	
		(CAL80836) clone JG35-K4-AG-narG1		
		Respiratory nitrate reductase, alpha subunit	144/204	
		[Geobacillus thermodenitrificans NG80-2]	(70.6%)	
CT30-narG_31	FN554861	dissimilatory membrane-bound nitrate	201/204	2
		reductase [uncultured bacterium]	(98.5%)	
		(CAL80836) clone JG35-K4-AG-narG1		
		Respiratory nitrate reductase, alpha subunit	143/204	
		[Geobacillus thermodenitrificans NG80-	(70.1%)	
		2] (YP_001124778)		
CT30-narG_44	FN554862	dissimilatory membrane-bound nitrate	204/205	2
		reductase [uncultured bacterium]	(99.5%)	
		(CAL80836) clone JG35-K4-AG-narG1		
		Respiratory nitrate reductase, alpha subunit	145/205	
		[Geobacillus thermodenitrificans NG80-2]	(70.7%)	
		(YP_001124778)		
CT30-narG_49	FN554863	membrane bound nitrate reductase	201/207	2
		[uncultured bacterium]	(97.1%)	
		(ABF20967) clone LT-075_21		
		Respiratory nitrate reductase alpha subunit	146/208	
		[Geobacillus thermodenitrificans NG80-2]	(70.2%)	
		(YP_001125821)		

Table 14. Quantities of *Geobacteracea* genomic DNA detected by real-time PCR with *Geobacteraceaea* specific primers (Geo564f and Geo840r) and qPCR SYBR Green Master Mix (Stratagene).

Well name	Well	Assay	Well type	Treshold	Ct	Quantity	RSq	Slope
	comment			(dR)	(dR)	(ng)	(dR)	(dR)
NTC	Negative	SYBR	NTC	471.392	No Ct	No Ct	0.98	-3.32
	control							
NPC	Primer control	SYBR	NPC	471.392	No Ct	No Ct	0.98	-3.32
G.s 1in5	G. sul 1 in 5	SYBR	Standard	471.392	17.9	2.700	0.98	-3.32
G.s 1 in 25	G. sul 1 in 25	SYBR	Standard	471.392	19.37	0.540	0.98	-3.32
G.s 1 in 125	G. sul 1 in 125	SYBR	Standard	471.392	21.86	0.108	0.98	-3.32
G. s 1 in 625	G. sul 1 in 625	SYBR	Standard	471.392	24.8	0.022	0.98	-3.32
A TO	A TO	SYBR	Unknown	471.392	No Ct	No Ct	0.98	-3.32
A T24	A T24	SYBR	Unknown	471.392	24.99	0.015	0.98	-3.32
C T0	C TO	SYBR	Unknown	471.392	No Ct	No Ct	0.98	-3.32
C T30	C T30	SYBR	Unknown	471.392	No Ct	No Ct	0.98	-3.32
C T60	C T60	SYBR	Unknown	471.392	No Ct	No Ct	0.98	-3.32
C T80	C T80	SYBR	Unknown	471.392	22.57	0.080	0.98	-3.32

1.6. References

- Binsted, N. CCLRC Daresbury Laboratory EXCURV98 Program, **1998**, Warrington, UK: CCLRC Daresbury Laboratory.
- Binsted, N; Strange, R.W.; Hasnain, S.S. Constrained and restrained refinement in EXAFS data analysis with curved wave theory. Biochemistry, **1992**, *31*, 12117–12125.
- Brewer, P. G.; Spencer, D. W. Colourimetric determination of Mn in anoxic waters. Limnol. Oceanogr. **1971**, *16*, 107-110.
- Gurman, S.J.; Binsted, N.; Ross, I. A rapid, exact curved-wave theory for EXAFS calculations. J. Physics C: Solid St Phy. **1984**, *17*,143–151.
- Harris, S. J.; Mortimer, R. J. G. Determination of nitrate in small water samples (100 μ L) by the cadmium-copper reduction method: A manual technique with application to the interstitial waters of marine sediments. Int. J. Environ. Anal. Chem. **2002**, *82*, 369-376.
- Lukens Jr., W.W.; Bucher, J.J.; Edelstein, N.M.; Shuh, D.K. Products of pertechnetate radiolysis in highly alkaline solution: structure of TcO₂-xH₂O. Environ. Sci. Technol. 2000, 36, 1124–1129.
- Maes, A.; Geraedts, K.; Bruggeman, C.; Vancluysen, J.; Rossberg, A.; Hennig, C. Evidence for the interaction of technetium colloids with humic substances by X-ray absorption spectroscopy. Environ. Sci. Technol. **2004**, *38*, 2044–2051.
- Morris, K.; Livens F.R.; Charnock J.M.; Burke I.T.; McBeth J.M.; Begg J.D.; Boothman C.; Lloyd J.R. An X-ray absorption study of the fate of technetium in reduced and reoxidised sediments and mineral phases, App. Geochem. **2008**, *23*, 603-617.
- Stookey L.L. Ferrozine a new spectrophotometric reagent for iron. Anal. Chem. **1970**, *42*, 779–781.
- Verardo, D. J., Froelich, P. N., and McIntyre, A. Determination of organic carbon and nitrogen in marine sediments using the Carlo Erba NA-1500 Analyser. Deep-Sea Res. 1990, 37, 157-165.
- Viollier, E.; Inglett, P. W.; Hunter, K.; Roychoudhury, P.; van Cappellen, P. The ferrozine method revisited: Fe(II)/Fe(III) determination in natural waters. Appl. Geochem. 2000, 15, 785-790.
- Wharton, M.J., Atkins, B.; Charnock, J.M.; Livens, F.R.; Pattrick, R.A.D.; Collison, D. An X-ray absorption spectroscopy study of the coprecipitation of Tc and Re with mackinawite (FeS). App. Geochem. **2000**, *15*, 347–354.

Wilkins, M.J.; Livens, F.R.; Vaughan, D.J.;Beadle, I.; Lloyd, J.R. The influence of microbial redox cycling on radionuclide mobility in the subsurface at a low-level radioactive waste storage site. Geobiology, **2007**, *5*, 293-301.